

# Blood film staining

In order to be examined microscopically all smears need to be stained. The same stain can be used in your clinic for manually staining blood films and cytology smears. For manual staining it is recommended that slides be immersed in stain-filled jars rather than covering slides with staining solution because formation of precipitate by evaporation can occur.

## Materials required:

Microscope slides\* / Solution 1 (methanol)\* / Solution 2 (eosinate)\* / Solution 3 (polychrome)\* / stain jars or containers.

*\* Can be obtained from Awanui Veterinary using online ordering.*

## How to:

1. Prepare a thin smear and air dry.
2. Dip the slide in Solution 1 (methanol) for 15 seconds to fix.
3. Allow excess methanol to drain off the slide.
4. Dip the slide in Solution 2 (eosinate) solution for 15 seconds.
5. Rinse the slide in water or else allow excess eosinate solution to drain off the slide.
6. Dip the slide in Solution 3 (polychrome) for 30 seconds.
7. Rinse the slide in water.
8. Air dry slide or dry using a hair drier – DO NOT blot.

## Lab tips:

- For storing staining solutions, use three slide-holder containers - one for each stain.
- Mark the containers with the date they were filled so that you know when to change them.
- The slides fit perfectly in the slots, so you will never again lose a slide in a stain container.
- Close the containers when not in use to prevent evaporation.

